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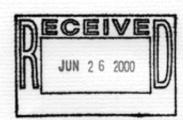
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Dr. C. W. Jameson National Toxicology Program Report on Carcinogens, MD EC-14 P.O.Box 12233 Research Triangle Park, NC 27709

June 20, 2000

Dear Dr. Jameson:



The American Academy of Dermatology [hereinafter referred to as "the Academy] submits these comments in response to the National Toxicology Program's recommendations to list ultraviolet radiation (UVR), nickel and nickel compounds, and human papillomaviruses (HPVs) in its tenth edition of the "Report on Carcinogens." The Academy supports the NTP's recommendations of inclusion of these materials on its list of known or suspected carcinogens.

Ultraviolet Radiation

Solar exposure is an environmental issue with profound effects for the majority of Americans. In this century, changes in attitudes of most Americans toward fashion and beauty as well as an increase in leisure activity outdoors has cost us dearly in terms of photodamage, photoaging, and photocarcinogenesis. In the last decade, however, public health education programs have been initiated to try to convince the American public of the error of its ways. This has been a public/private partnership with such entities as the Environmental Protection Agency, the National Institutes of Health, the Centers for Disease Control and Prevention, the American Academy of Dermatology, the American Cancer Society, The Skin Cancer Foundation and others providing educational materials in a variety of formats and media. We believe that adding UVR to the list of known carcinogens would be of great importance to our efforts to convince the public that unprotected sun exposure is dangerous and may lead to the development of melanoma and non-melanoma skin cancers, such as basal cell and squamous cell carcinoma.

This year, the American Academy of Dermatology estimates that 1.3 million Americans will be diagnosed with some form of skin cancer. The causal agent for the majority of these skin cancers is exposure to UVR. Genetic damage from UVR occurs in two general forms, mutations and chromosome damage. Both are important to the development of the majority of neoplasms seen on the skin - both benign and malignant lesions. Exposure to UVR leads to mutations in the DNA by at least two potential mechanisms. In general, mutations arise in the skin when UV-induced covalent damage is misrepaired, altering the base sequence from the original. Alternatively, undetected covalent damage that is not repaired prior to DNA replication could induce a misread by DNA polymerase, causing a base substitution that alters the original sequence.

Pyrimidine dimers and 6-4 photoproducts are two of the most important classes of direct DNA damage induced by UV. The peak wavelength for the formation of pyrimidine dimers and 6-4 photoproducts is 300 nm, so it is more than likely that UVB radiation is the major source of mutations in the human skin. In addition, most of the mutations seen in the p53 gene, an important tumor suppressor gene in human skin cancer are C→T and C→TT transitions. This is consistent with UVB-induced pyrimidine dimer formation as the initial effect. While UVB radiation can induce DNA damage directly through the formation of photoproducts, *in vitro* studies have shown that UVA requires an intermediate molecular target to generate the reactive oxygen species that are thought to be the primary mediators of UVA-induced damage.

UVB and UVA radiation can induce DNA damage indirectly, through the generation of reactive oxygen species, also known as free radicals. Products such as superoxide, singlet oxygen, hydrogen peroxide and hydroxyl radials – all generate from UV striking molecular targets in cells – can introduce DNA adducts or other covalent changes in the DNA. When covalent changes are unrecognized or unrepaired, they may be misread at the time of DNA replication prior to cell division, leading to permanent mutations in the DNA base sequences.

A characteristic change in DNA due to oxidative effects of UV radiation is the formation of 8-hydroxyguanine, a derivative of the purine base deoxyguanine. Base modifications characteristic of oxidative damage are produced by wavelengths throughout the UVA and UVB spectra, although the yield of pyrimidine dimers decreases exponentially above 315 nm – near the transition from UVB to UVA. In addition, there is a second peak of oxidative damage that occurs in the visible range between 400 and 450 nm. This suggests that blue light may also contribute to DNA damage. Support for the possibility that visible light may contribute to carcinogenesis comes from a fish model of melanoma, in which wavelengths in this range can trigger tumor development. More research, however, is needed in this area.

UV-induced chromosome damage is less well understood than UV-induced mutations, although there is abundant evidence that this is a major pathway for tumorigenesis. Chromosome translocations can inactivate tumor suppressor genes or up-regulate tumorigenic oncogenes. Non-random chromosome deletions are an important means of inactivation of function for tumor suppression genes, as control gene function is lost when a germ line defect combines with a somatic event or when two somatic events occur in both alleles.

In addition to skin cancers, there is strong epidemiological evidence linking UVR exposure to other malignancies. Data from the Swedish cancer registry indicates that UVR exposure may be an etiologic factor in non-Hodgkin's lymphoma. Much more research needs to be done in this area, but if these epidemiological observations are true, UVR may be a far more significant carcinogen than is currently appreciated.

Exposure to UVR has other deleterious health effects. Many of the cutaneous changes that we associate with aging are due primarily to exposure to UVR, and therefore are potentially preventable. Animal models have shown that both UVA and UVB contribute to photoaging. These cutaneous changes are caused by oxidative damage and stress, such as lipid peroxidation, as well as erythema. UVR effect on cytokine release and signal transduction pathways may alter the expression of enzymes that remodel the dermis. A photoaging-related effect of chronic exposure to UVR is solar elastosis, or the presence of abnormal elastin in the skin. Metalloproteinases, such as collagenase, can be up-regulated by UVB exposures equivalent to 1/6 of the minimal dose of UVB that produces erythema; this effect contributes to the development of cutaneous photoaging.

There are also a number of photosensitive disorders in which UVR plays a significant role. The two most studied are polymorphous light eruption (PMLE) and lupus erythematosis. PMLE is characterized by pruritic inflammatory skin eruptions seen primarily on the arms and upper trunk. PMLE is a relatively common disorder, affecting 10% of the general population. Individuals of Native American and Scandinavian ancestry are more at risk for these abnormal reactions to sunlight.

Lupus erythematosis is an autoimmune disorder characterized by the presence of inappropriate antibodies to self-antigens. Autoantibodies to DNA are highly characteristic of the disease. As with PMLE, UVB and UVA and perhaps visible light can play a role in the induction of lupus. This suggests that oxidative damage may play an important role in the disorder. Photosensitivity is a common manifestation of several forms of lupus, and tends to correlate with a less favorable prognosis and more organ involvement in systemic lupus and exacerbations of cutaneous lupus.

Nickel and Nickel Compounds

The Academy joins with our colleagues in the American Contact Dermatitis Society in supporting efforts that would reduce human exposure to nickel and nickel compounds. We ask that the NTP follow the lead of our European counterparts to reduce occupational and consumer exposure to nickel.

Currently, human exposure to nickel is common, as it is used widely in both industry and in consumer products. Experimental and epidemiological data have shown that sparingly soluble nickel compounds, and possibly also the soluble compounds, are carcinogenic in humans. Exposure to these metals has been linked to the development of lung and nasal cancers. The presumed route of exposure for carcinogenesis has been inhalation, although recently exposures from medical and dental devices have also been scrutinized. Furthermore, it has been hypothesized that certain paternal exposures to nickel may

increase the risk of cancer in progeny. The mechanism by which nickel induces carcinogenicity, however, still remains unclear, but may be caused by direct or indirect actions of nickel compounds on DNA, co-carcinogenicity by deregulating cellular proliferation, and/or tumor promotion. Much more research needs to be undertaken to determine which compounds are co-carcinogens and which act as tumor promoters.

In addition to its possible links to cancer, exposure to nickel causes another health effect of considerable morbidity – allergic contact dermatitis. Twenty years ago, epidemiological evidence showed that nearly 10% of the US population exhibited some sensitivity to nickel. Since that time, the incidence of nickel allergy has increased dramatically. In a recent study published in Norway, approximately 30% of women in two different regions of that country were found to be allergic to nickel, while the incidence rate among men was approximately 5%. Scientists postulate that gender differences in the incidence rates of nickel allergy maybe due primarily to the common practice of body piercing.

Because of the increasing rate of nickel sensitization, the Danish Ministry of the Environment recently issued a statutory order limiting the permissible release of nickel from objects intended for close contact with the skin to $\leq 0.5~\mu g/cm^2$. These items include earrings, eyeglass frames, and buttons. This level was based on a number of studies that indicated the relative lack of sensitization to nickel at concentrations at or below $0.5~\mu g/cm^2$ per week. Since the adoption of these nickel restrictions, the frequency of nickel allergy among children decreased from a high of 24.8% prior to the enactment of the new standard to 9.2%.

Shortly, the European Community will enact the Directives of the European Standards for the Analytical Methods to be used on the nickel directive. This directive states that objects, which come into direct and prolonged contact with the skin, must not contain more than 0.5 µg/cm² of nickel. Although this new law will prevent new cases of nickel sensitization, it will unfortunately have little effect on those individuals already sensitive to the metal. However, given the growing incidence of nickel allergy, prevention is very important and I would urge that the NTP consider restricting nickel exposure through the adoption of the European standard.

Human Papillomaviruses

The Academy supports the listing Human papillomavirus (HPV) as a known human carcinogen. Historically, dermatologists have played a crucial role in the diagnosis and treatment of sexually transmitted diseases (STDs), because many of these infections present predominantly with cutaneous signs and symptoms. Given our experience in vast experience in treating this prevalent viral STD, it is our expert opinion that listing HPV as a known carcinogen will assist us in our efforts to educate the general public about the dangers of this disease, how it may be prevented, how it is diagnosed, and how it is treated.

There are over 70 distinct types of HPV. Nearly half, 30 types, are transmitted sexually by skin-to-skin contact and cause genital HPV. According to the American Social Health Association, 5.5 million new cases of sexually transmitted HPV occur each year, and 20 million Americans are thought to have an active HPV infection at any given time. This year, direct annual medical costs for treating the symptoms of HPV infection is expected to reach \$6 billion.

While many forms of the virus are harmless, certain strains of HPV have been clearly linked to the development of cervical lesions and then to cervical cancers. Indeed, over 99% of cervical cancers are associated with HPV. Cervical cancer is the second most common cacer of women in the world. In 1999, 500,000 women in the United States were diagnosed with cervical cancer, and 200,000 died from it. As with skin cancers, the lag time between exposure and the development of the cervical cancer is often lengthy, sometimes between 10 and 20 years. In addition to cervical cancers, exposure to certain strains of HPV is linked to the development of other cancers such as carcinomas of the nasal septum, laryngeal and hypopharyngeal carcinoma, cancers of the upper digestive and respiratory tract, as well as other anogenital cancers.

Unfortunately, the majority of Americans are unaware of the linkage between certain strains of HPV and cervical and other cancers. According to an expert panel convened in 1999 by the Centers for Disease Control and Prevention, many health care providers were equally unaware of the risks of exposure to HPV and the development of cervical cancer as well as to the new screening techniques for HPV and treatment modalities for cervical cancer.

Listing HPV as a known carcinogen will bring a clarity to our public health messages concerning HPV. It will provide an impetus to clinicians to learn more about the dangers of this viral STD and will encourage those at risk to be tested.

In summary, the Academy supports the listing of UVR, nickel and nickel compounds, and HPV to the NTP's tenth edition of the "Report on Carcinogens." A comprehensive list of citations from peer-reviewed clinical journals and Academy reports is appended to this letter. If I can be of further assistance to you in your deliberations, please do not hesitate to contact me.

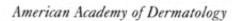
Sincerely,

President

Richard Scher, M.D.

**PLEASE NOTE: Human Papillomavirus is being deferred for review until the 2001 review cycle.

Report on Carcinogens Group, NIEHS/NTP



Citations - UV Radiation

References

- Runger TM, Epe B, Moller K. Processing of directly and indirectly ultraviolet-induced DNA damage in human cells. Recent Results in Cancer Research. 1995;139:31-42.
- Tornaletti S, Rozek D, Pfeifer GP. The distribution of UV photoproducts along the human p53 gene and its relation to mutations in skin cancer. Oncogene. 1993;8:2051-2057.
- Freeman SE, Hacham H, Gange RW, et al. Wavelength dependence of pyrimidine dimer formation in DNA of human skin irradiated in situ with ultraviolet light. Proc Natl Acad Sci. 1989;86:5605-5609.
- DeMarini DM, Shelton ML, Stankowski LF, Jr. Mutation spectra in Salmonella of sunlight, white fluorescent light, and light from tanning salon beds: induction of tandem mutations and role of DNA repair. *Mutation Res*. 1995;327:131-49.
- Drouin R, Therrien JP. UVB-induced cyclobutane pyrimidine dimer frequency correlates with skin cancer mutational hotspots in p53. Photochem Photobiol. 1997;66:719-26.
- McGregor JM, Crook T, Fraser-Andrews EA, et al. Spectrum of p53 gene mutations suggests a possible role for ultraviolet radiation in the pathogenesis of advanced cutaneous lymphomas. J Invest Dermatol. 1999;112:317-21.
- Pierceall WE, Mukhopadhyay T, Goldberg LH, Ananthaswamy HN.
 Mutations in the p53 tumor suppressor gene in human cutaneous squamous cell carcinomas. Mol Carcinogenesis. 1991;4:445-9.
- Zhang X, Rosenstein B, Wang Y, Lebwohl M, Wei H. Identification of possible reactive oxygen species involved in ultraviolet radiation-induced oxidative DNA damage. Free Radic Biol Med. 1997;23:980-985.
- Kielbassa C, Roza L, Epe B. Wavelength dependence of oxidative DNA damage induced by UV and visible light. Carcinogenesis. 1997;18:811-6.
- Setlow RB, Grist E, Thompson K, Woodhead AD. Wavelengths effective in induction of malignant melanoma. Proc Natl Acad Sci. 1993;90:6666-6670.
- Zhang X, Rosenstein BS, Wang Y, et al. Induction of 8-oxo-7,8-dihydro-2'deoxyguanosine by ultraviolet radiation in calf thymus DNA and HeLa cells. Photochem and Photobiol. 1997;65:119-124.

- Rosen JE, Prahalad AK, Williams GM. 8-Oxodeoxyguanosine formation in the DNA of cultured cells after exposure to H2O2 alone or with UVB or UVA irradiation [published erratum appears in Photochem Photobiol 1996 Sep;64(3):611]. Photochemistry & Photobiology. 1996;64:117-22.
- Richmond A, Fine R, Murray D, Lawson D. Growth factor and cytogenetic abnormalities in cultured nevi and malignant melanomas. J Invest Dermatol. 1986;86:295-302.
- Parmiter AH, Nowell PC. Cytogenetics of melanocytic tumors. J Invest Dermatol. 1993;100:254S-258S.
- Albino AP, Fountain JW. Molecular genetics of human malignant melanoma. Cancer Treatment and Research. 1993;65:201-255.
- Becher R, Gibas Z, Karakousis C, Sandberg AA. Nonrandom chromosome changes in malignant melanoma. Cancer Res. 1983;43:5010-5016.
- Trent JM, Leong SP, Meyskens FL. Chromosome alterations in human malignant melanoma. Carcinog Compr Surv. 1989;11:165-186.
- Trent JM. Cytogenetics of human malignant melanoma. Cancer & Metastasis Reviews. 1991;10:103-113.
- Wiltshire RN, Duray P, Bittner ML, et al. Direct visualization of the clonal progression of primary cutaneous melanoma: application of tissue microdissection and comparative genomic hybridization. Cancer Res. 1995;55:3954-3957.
- Pedersen MI, Wang N. Chromosomal evolution in the progression and metastasis of human malignant melanoma. A multiple lesion study. Cancer Genetics & Cytogenetics. 1989;41:185-201.
- Ochi H, Wake N, Rao U, et al. Serial cytogenetic analysis of a recurrent malignant melanoma. Cancer Genetics & Cytogenetics. 1984;11:175-183.
- Peris K, Keller G, Chimenti S, et al. Microsatellite instability and loss of heterozygosity in melanoma. J Invest Dermatol. 1995;105:625-628.
- Healy E, Rehman I, Angus B, Rees J. Loss of heterozygosity in sporadic primary cutaneous melanoma. Genes, Chromosomes & Cancer. 1995;12:152-156.
- Naylor MF, Brown S, Quinlan C, Pitha JV, Everett MA. 9p21 deletions in primary melanoma. Dermatology Online Journal. 1997;3:1.

- Dreosti I, McGown M. Antioxidants and UV-induced genotoxicity. Research Communications in Chemical Pathology & Pharmacology. 1992;75:251-4.
- Harrison JA, Walker SL, Plastow SR, et al. Sunscreens with low sun protection factor inhibit ultraviolet B and A photoaging in the skin of the hairless albino mouse. Photodermatol Photoimmunol Photomed. 1991;8:12-20.
- Kligman LH. The ultraviolet-irradiated hairless mouse: a model for photoaging. J Amer Acad Dermatol. 1989;21:623-631.
- 28. Fisher GJ, Datta SC, Talwar HS, et al. Molecular basis of sun-induced premature skin ageing and retinoid antagonism. *Nature*. 1996;379:335-339.
- Gonzalez S, Pathak MA. Inhibition of ultraviolet-induced formation of reactive oxygen species, lipid peroxidation, erythema and skin photosensitization by polypodium leucotomos. *Photodermatol Photoimmunol Photomed*. 1996;12:45-56.
- Bissett DL, Chatterjee R, Hannon DP. Photoprotective effect of superoxide-scavenging antioxidants against ultraviolet radiation-induced chronic skin damage in the hairless mouse. *Photodermatol Photoimmunol Photomed*. 1990;7:56-62.
- Kawaguchi Y, Tanaka H, Okada T, et al. Effect of reactive oxygen species on the elastin mRNA expression in cultured human dermal fibroblasts. Free Radical Biology & Medicine. 1997;23:162-5.
- Kawaguchi Y, Tanaka H, Okada T, et al. The effects of ultraviolet A and reactive oxygen species on the mRNA expression of 72-kDa type IV collagenase and its tissue inhibitor in cultured human dermal fibroblasts. Arch Dermatol Res. 1996;288:39-44.
- Fisher GJ, Voorhees JJ. Molecular mechanisms of retinoid action in skin. FASEB J. 1996;10:1002-1013.
- Kang S, Fisher GJ, Voorhees JJ. Photoaging and topical tretinoin: therapy, pathogenesis, and prevention. Arch Dermatol. 1997;133:1280-1284.
- Grant AJ, Jessup W, Dean RT. Accelerated endocytosis and incomplete catabolism of radical-damaged protein. Biochem Biophys Acta. 1992;1134:203-9.
- Dean RT, Fu S, Stocker R, Davies MJ. Biochemistry and pathology of radical-mediated protein oxidation. *Biochem J*. 1997;324:1-18.

- Wagenmakers AJ, Reinders RJ, van Venrooij WJ. Cross-linking of mRNA to proteins by irradiation of intact cells with ultraviolet light. Eur J Biochem. 1980;112:323-30.
- 38. Davies KJ, Delsignore ME. Protein damage and degradation by oxygen radicals. III. Modification of secondary and tertiary structure. *J Biol Chem.* 1987;262:9908-13.
- Davies KJ, Lin SW, Pacifici RE. Protein damage and degradation by oxygen radicals. IV. Degradation of denatured protein. J Biol Chem. 1987;262:9914-20.
- Kligman LH, Akin FJ, Kligman AM. Sunscreens promote repair of ultraviolet radiation-induced dermal damage. J Invest Dermatol. 1983;81:98-102.
- 41. Kligman LH, Akin FJ, Kligman AM. Prevention of ultraviolet damage to the dermis of hairless mice by sunscreens. *J Invest Dermatol.* 1982;78:181-189.
- Jensen P, Hansen S, Moller B, et al. Skin cancer in kidney and heart transplant recipients and different long-term immunosuppressive therapy regimens. J Am Acad Dermatol. 1999;40:177-86.
- Ong CS, Keogh AM, Kossard S, Macdonald PS, Spratt PM. Skin cancer in Australian heart transplant recipients. J Am Acad Dermatol. 1999;40:27-34.
- Kripke ML, Fisher MS. Immunologic parameters of ultraviolet carcinogenesis. J Natl Cancer Inst. 1976;57:211-215.
- Noonan FP, De F, E.C., Kripke ML. Suppression of contact hypersensitivity by UV radiation and its relationship to UV-induced suppression of tumor immunity. Photochemistry & Photobiology. 1981;34:683-689.
- Elmets CA, Bergstresser PR, Tigelaar RE, Wood PJ, Streilein JW.
 Analysis of the mechanism of unresponsiveness produced by haptens painted on skin exposed to low dose ultraviolet radiation. J Exp Med. 1983;158:781-794.
- Okamoto H, Kripke ML. Effector and suppressor circuits of the immune response are affected in vivo by different mechanisms. Proc Natl Acad Sci. 1987;84:3841-3845.
- Nishigori C, Yarosh DB, Donawho C, Kripke ML. The immune system in ultraviolet carcinogenesis. *Journal of Investigative Dermatology Symposium Proceedings*. 1996;1:143-146.

- 49. Kripke ML, Cox PA, Alas LG, Yarosh DB. Pyrimidine dimers in DNA initiate systemic immunosuppression in UV- irradiated mice. *Proc Natl Acad Sci U S A*. 1992;89:7516-20.
- Applegate LA, Ley RD, Alcalay J, Kripke ML. Identification of the molecular target for the suppression of contact hypersensitivity by ultraviolet radiation. J Exp Med. 1989;170:1117-1131.
- 51. O'Connor A, Nishigori C, Yarosh D, et al. DNA double strand breaks in epidermal cells cause immune suppression in vivo and cytokine production in vitro. *J Immunol.* 1996;157:271-8.
- Kripke ML, Cox PA, Bucana C, et al. Role of DNA damage in local suppression of contact hypersensitivity in mice by UV radiation. Exp Dermatol. 1996;5:173-80.
- Garssen J, Vandebriel RJ, van Loveren H. Molecular aspects of UVBinduced immunosuppression. Arch Toxicol Suppl. 1997;19:97-109.
- Toews GB, Bergstresser PR, Streilein JW. Epidermal langerhans cell density determines whether contact hypersensitivity or unresponsiveness follows skin painting with DNFB. J Immunol. 1980;124:445-453.
- Wolf P, Cox P, Yarosh DB, Kripke ML. Sunscreens and T4N5 Liposomes differ in their ability to protect against ultraviolet-induced sunburn cell formation, alterations of dendritic epidermal cells, and local suppression of contact hypersensitivity. J Invest Dermatol. 1995;104:287-292.
- Cooper KD, Duraiswamy N, Hammerberg C, et al. Neutrophils, differentiated macrophages, and monocyte/macrophage antigen presenting cells infiltrate murine epidermis after UV injury. J Invest Dermatol. 1993;101:155-163.
- Rivas JM, Ullrich SE. Systemic suppression of delayed-type hypersensitivity by supernatants from UV-irradiated keratinocytes. An essential role for keratinocyte-derived IL-10. *J Immunol.* 1992;149:3865-3871.
- Rivas JM, Ullrich SE. The role of IL-4, IL-10, and TNF-alpha in the immune suppression induced by ultraviolet radiation. *Journal of Leukocyte Biology*. 1994;56:769-775.
- Kurimoto I, Streilein JW. Deleterious effects of cis-urocanic acid and UVB radiation on Langerhans cells and on induction of contact hypersensitivity are mediated by tumor necrosis factor-alpha. J Invest Dermatol. 1992;99:69S-70S.
- Robertson B, Gahring L, Newton R, Daynes R. In vivo administration of interleukin 1 to normal mice depresses their capacity to elicit contact

hypersensitivity responses: prostaglandins are involved in this modification of immune function. J Invest Dermatol. 1987;88:380-387.

- Miyauchi-Hashimoto H, Okamoto H, Tanaka K, Horio T. Ultraviolet radiation-induced suppression of natural killer cell activity is enhanced in xeroderma pigmentosum group A (XPA) model mice. J Invest Dermatol. 1999;112:965-70.
- De Fabo EC, Noonan FP. Mechanism of immune suppression by ultraviolet irradiation in vivo. I. Evidence for the existence of a unique photoreceptor in skin and its role in photoimmunology. J Exp Med. 1983;158:84-98.
- Bestak R, Barnetson RS, Nearn MR, Halliday GM. Sunscreen protection of contact hypersensitivity responses from chronic solar-simulated ultraviolet irradiation correlates with the absorption spectrum of the sunscreen. J Invest Dermatol. 1995;105:345-351.
- Damian DL, Halliday GM, Barnetson RS. Broad-spectrum sunscreens provide greater protection against ultraviolet-radiation-induced suppression of contact hypersensitivity to a recall antigen in humans. J Invest Dermatol. 1997;109:146-151.
- Moyal D. Immunosuppression induced by chronic ultraviolet irradiation in humans and its prevention by sunscreens. Eur J Dermatol. 1998;8:209-11.
- Strickland FM, Pelley RP, Kripke ML. Prevention of ultraviolet radiationinduced suppression of contact and delayed hypersensitivity by Aloe barbadensis gel extract. J Invest Dermatol. 1994;102:197-204.
- Atillasoy ES, Elenitsas R, Sauter ER, Soballe PW, Herlyn M. UVB induction of epithelial tumors in human skin using a RAG-1 mouse xenograft model. J Invest Dermatol. 1997;109:704-9.
- Green AC, Siskind V, Bain C, Alexander J. Sunburn and malignant melanoma. Br J Cancer. 1985;51:393-397.
- Whiteman D, Green A. Melanoma and sunburn. Cancer Causes & Control. 1994;5:564-572.
- MacKie RM, Aitchison T. Severe sunburn and subsequent risk of primary cutaneous malignant melanoma in Scotland. Br J Cancer. 1982;40:955-960.
- Grodstein F, Speizer FE, Hunter DJ. A prospective study of incident squamous cell carcinoma of the skin in the nurses' health study. J Natl Cancer Inst. 1995;87:1061-1066.

- 72. Kricker A, Armstrong BK, English DR, Heenan PJ. Does intermittent sun exposure cause basal cell carcinoma? a case-control study in Western Australia. *Int J Cancer.* 1995;60:489-494.
- Gallagher RP, Hill GB, Bajdik CD, et al. Sunlight exposure, pigmentation factors, and risk of nonmelanocytic skin cancer. II. Squamous cell carcinoma. Arch Dermatol. 1995;131:164-169.
- Green A, Williams G, Neale R, et al. Daily sunscreen application and betacarotene supplementation in prevention of basal-cell and squamous-cell carcinomas of the skin: a randomized controlled trial. Lancet. 1999;354:723-9.
- 75. Eller MS, Yaar M, Gilchrest BA. DNA damage and melanogenesis. *Nature*. 1994;372:413-414.
- 76. Gilchrest BA, Park HY, Eller MS, Yaar M. Mechanisms of ultraviolet light-induced pigmentation. *Photochemistry & Photobiology*. 1996;63:1-10.
- Green AC, O'Rourke MG. Cutaneous malignant melanoma in association with other skin cancers. J Natl Cancer Inst. 1985;74:977-80.
- 78. Crombie IK. Variation of melanoma incidence with latitude in North America and Europe. *Br J Cancer*. 1979;40:774-781.
- Eklund G, Malec E. Sunlight and incidence of cutaneous malignant melanoma. Effect of latitude and domicile in Sweden. Scandinavian Journal of Plastic & Reconstructive Surgery. 1978;12:231-241.
- Elwood JM, Lee JA, Walter SD, Mo T, Green AE. Relationship of melanoma and other skin cancer mortality to latitude and ultraviolet radiation in the United States and Canada. *International Journal of Epidemiology*. 1974;3:325-332.
- Bulliard JL, Cox B, Elwood JM. Latitude gradients in melanoma incidence and mortality in the non-Maori population of New Zealand. Cancer Causes & Control. 1994;5:234-40.
- Lee JA, Scotto J. Melanoma: linked temporal and latitude changes in the United States. Cancer Causes & Control. 1993;4:413-8.
- Beitner H, Norell SE, Ringborg U, Wennersten G, Mattson B. Malignant melanoma: aetiological importance of individual pigmentation and sunexposure. Br J Cancer. 1990;122:43-51.
- 84. Rodenas JM, Delgado-Rodriguez M, Herranz MT, Tercedor J, Serrano S. Sun exposure, pigmentary traits, and risk of cutaneous malignant melanoma: a

case-control study in a Mediterranean population. Cancer Causes & Control. 1996;7:275-83.

- Kobayashi N, Muramatsu T, Yamashina Y, et al. Melanin reduces ultraviolet-induced DNA damage formation and killing rate in cultured human melanoma cells. J Invest Dermatol. 1993;101:685-689.
- 86. Garbe C, Buttner P, Weiss J, et al. Risk factors for developing cutaneous melanoma and criteria for identifying persons at risk: multicenter case-control study of the Central Malignant Melanoma Registry of the German Dermatological Society. *J Invest Dermatol.* 1994;102:695-699.
- Crombie IK. Racial differences in melanoma incidence. Br J Cancer. 1979;40:185-193.
- Bell M, Beyl CM, Schopf RE, Schramm P. Light exposure of the lower leg as a pathogenetic factor in the occurrence of malignant melanoma. *Dermatology*. 1992;185:257-61.
- Cress RD, Holly EA, Ahn DK, LeBoit PE, Sagebiel RW. Cutaneous melanoma in women: anatomic distribution in relation to sun exposure and phenotype. Cancer Epidemiol Biomarkers Prev. 1995;4:831-6.
- 90. Crombie IK. Distribution of malignant melanoma on the body surface area. Br J Cancer. 1981;43:842-849.
- Atillasoy ES, Seykora JT, Soballe PW, et al. UVB induces atypical melanocytic lesions and melanoma in human skin. Am J Pathol. 1998;152:1179-1186.
- Kusewitt DF, Applegate LA, Ley RD. Ultraviolet radiation-induced skin tumors in a South American opossum (Monodelphis domestica). Vet Pathol. 1991;28:55-65.
- Setlow RB, Wookhead AD, Grist E. Animal model for ultraviolet radiationinduced melanoma: platyfish-swordtail hybrid. Proc Natl Acad Sci. 1989;86:8922-8926.
- 94. de Gruijl FR. Action spectrum for photocarcinogenesis. Recent Results in Cancer Research. 1995;139:21-30.
- de Laat A, van der Leun JC, de Gruijl FR. Carcinogenesis induced by UVA (365-nm) radiation: the dose-time dependence of tumor formation in hairless mice. Carcinogenesis. 1997;18:1013-1020.

- Bech-Thomsen N, Wulf HC, Poulsen T, Christensen FG, Lundgren K.
 Photocarcinogenesis in hairless mice induced by ultraviolet A tanning devices with or without subsequent solar-simulated ultraviolet irradiation. *Photodermatol Photoimmunol Photomed*. 1991;8:139-145.
- Matsui MS, DeLeo VA. Longwave ultraviolet radiation and promotion of skin cancer. Cancer Cells. 1991;3:8-11.
- Talve L, Stenback F, Jansen CT. UVA irradiation increases the incidence of epithelial tumors in UVB-irradiated hairless mice. *Photodermatol Photoimmunol Photomed*. 1990;7:109-115.
- Campbell C, Quinn AG, Angus B, Farr PM, Rees JL. Wavelength specific patterns of p53 induction in human skin following exposure to UV radiation. Cancer Res. 1993;53:2697-2699.
- 100. Schiemann WP, Pfeifer WM, Levi E, Kadin ME, Lodish HF. A deletion in the gene for transforming growth factor beta type I receptor abolishes growth regulation by transforming growth factor beta in a cutaneous T-cell lymphoma. *Blood.* 1999;94:2854-61.
- 101. Karenko L, Kahkonen M, Hyytinen ER, Lindlof M, Ranki A. Notable losses at specific regions of chromosomes 10q and 13q in the Sezary syndrome detected by comparative genomic hybridization [letter]. J Invest Dermatol. 1999;112:392-5.
- Berg M. Epidemiological studies of the influence of sunlight on the skin. Photodermatol. 1989;6:80-4.
- Morison WL, Stern RS. Polymorphous light eruption: a common reaction uncommonly recognized. Acta Derm Venereol. 1982;62:237-40.
- Epstein JH. Polymorphous light eruption. J Am Acad Dermatol. 1980;3:329-43.
- Hurwitz S, Clinical pediatric dermatology. 1st ed. 1981, Philadelphia: W.B. Saunders Co. 481.
- 106. Boonstra HE, van Weelden H, Toonstra J, van Vloten WA. Polymorphous light eruption: A clinical, photobiologic, and follow-up study of 110 patients. *J Am Acad Dermatol*. 2000;42:199-207.
- Lambert J, Verheyen A, Dockx P. Experimental reproduction of polymorphous light eruption and benign summer light eruption by whole-body UVA irradiation. *Dermatology*. 1997;194:388-91.

- 108. Ortel B, Tanew A, Wolff K, Honigsmann H. Polymorphous light eruption: action spectrum and photoprotection. *J Am Acad Dermatol.* 1986;14:748-53.
- 109. Moyal D, Binet O, Richard A, Rougier A, Hourseau C. Prevention of polymorphous light eruption by a new broadspectrum sunscreen: need for a high UVA protecting factor. in Annual Meeting of the American Academy of Dermatology. 1999, New Orleans, Louisiana USA:
- Morita A, Grewe M, Grether-Beck S, Olaizola-Horn S, Krutmann J.
 Induction of proinflammatory cytokines in human epidermoid carcinoma cells by in vitro ultraviolet A1 irradiation. *Photochem Photobiol*. 1997;65:630-5.
- Hadshiew I, Stab F, Untiedt S, et al. Effects of topically applied antioxidants in experimentally provoked polymorphous light eruption. Dermatology. 1997;195:362-8.
- 112. Thompson D, Juby A, Davis P. The clinical significance of autoantibody profiles in patients with systemic lupus erythematosus. *Lupus*. 1993;2:15-9.
- Blatt NB, Glick GD. Anti-DNA autoantibodies and systemic lupus erythematosus. Pharmacol Ther. 1999;83:125-39.
- Rahman MA, Isenberg DA. Autoantibodies in systemic lupus erythematosus. Curr Opin Rheumatol. 1994;6:468-73.
- Cooke MS, Mistry N, Wood C, Herbert KE, Lunec J. Immunogenicity of DNA damaged by reactive oxygen species—implications for anti-DNA antibodies in lupus. Free Radic Biol Med. 1997;22:151-9.
- 116. Blount S, Griffiths H, Emery P, Lunec J. Reactive oxygen species modify human DNA, eliciting a more discriminating antigen for the diagnosis of systemic lupus erythematosus. Clin Exp Immunol. 1990;81:384-9.
- Hasan T, Nyberg F, Stephansson E, et al. Photosensitivity in lupus erythematosus, UV photoprovocation results compared with history of photosensitivity and clinical findings. Br J Dermatol. 1997;136:699-705.
- Klein LR, Elmets CA, Callen JP. Photoexacerbation of cutaneous lupus erythematosus due to ultraviolet A emissions from a photocopier. Arthritis Rheum. 1995;38:1152-6.
- Lehmann P, Holzle E, Kind P, Goerz G, Plewig G. Experimental reproduction of skin lesions in lupus erythematosus by UVA and UVB radiation [see comments]. J Am Acad Dermatol. 1990;22:181-7.

- Nyberg F, Hasan T, Skoglund C, Stephansson E. Early events in ultraviolet light-induced skin lesions in lupus erythematosus: expression patterns of adhesion molecules ICAM-1, VCAM-1 and E-selectin. Acta Derm Venereol. 1999;79:431-6.
- van Weelden H, Velthuis PJ, Baart de la Faille H. Light-induced skin lesions in lupus erythematosus: photobiological studies [see comments]. Arch Dermatol Res. 1989;281:470-4.
- Velthuis PJ, van Weelden H, van Wichen D, Baart de la Faille H.
 Immunohistopathology of light-induced skin lesions in lupus erythematosus. Acta Derm Venereol. 1990;70:93-8.
- Sontheimer RD. Photoimmunology of lupus erythematosus and dermatomyositis: a speculative review. Photochem Photobiol. 1996;63:583-94.
- Tebbe B, Mansmann U, Wollina U, et al. Markers in cutaneous lupus erythematosus indicating systemic involvement. A multicenter study on 296 patients. Acta Derm Venereol. 1997;77:305-8.
- Doria A, Biasinutto C, Ghirardello A, et al. Photosensitivity in systemic lupus erythematosus: laboratory testing of ARA/ACR definition. Lupus. 1996;5:263-8.
- Vila LM, Mayor AM, Valentin AH, et al. Association of sunlight exposure and photoprotection measures with clinical outcome in systemic lupus erythematosus. P R Health Sci J. 1999;18:89-94.
- Adami J, Gridley G, Nyren O, et al. Sunlight and non-Hodgkin's lymphoma: a population-based cohort study in Sweden. Int J Cancer. 1999;80:641-5.
- Nole GE, Johnson AW, Cheney MC, Znaiden A. Cumulative lifetime UVR exposure in the United States and the effect of various levels of sunscreen protection. Cosmetic Dermatology. 1999;12:23-26.
- Kligman LH, Akin FJ, Kligman AM. Sunscreens prevent ultraviolet photocarcinogenesis. J Amer Acad Dermatol. 1980;3:30-35.
 Wulf HC, Poulsen T, Brodthagen H, Hou-Jensen K. Sunscreens for delay of ultraviolet induction of skin tumors. J Amer Acad Dermatol. 1982;7:194-202.
- Naylor MF, Boyd A, Smith DW, et al. High Sun Protection Factor (SPF) Sunscreens in the Suppression of Actinic Neoplasia. Arch Dermatol. 1995;131:170-175.

- Thompson SC, Jolley D, Marks R. Reduction of solar keratoses by regular sunscreen use. N Engl J Med. 1993;329:1147-1151.
- Setlow RB. Spectral regions contributing to melanoma: a personal view. J Investig Dermatol Symp Proc. 1999;4:46-9.
- 134. Gallagher CH, Greenoak GE, Reeve VE, et al. Ultraviolet carcinogenesis in the hairless mouse skin. Influence of the sunscreen 2-ethylhexyl-pmethoxycinnamate. Australian Journal of Experimental Biology & Medical Science. 1984;62:577-588.
- Naylor MF, Farmer KC. The Case for Sunscreens: a Review of Their Use in Preventing Actinic Damage and Neoplasia. Arch Dermatol. 1997;133:1146-1154.
- Knowland J, McKenzie EA, McHugh PJ, Cridland NA. Sunlight-induced mutagenicity of a common sunscreen ingredient. FEBS Letters. 1993;324:309-313.
- 137. McHugh PJ, Knowland J. Characterization of DNA damage inflicted by free radicals from a mutagenic sunscreen ingredient and its location using an in vitro genetic reversion assay. *Photochemistry & Photobiology*. 1997;66:276-81.
- Polyak K, Xia Y, Zweier J, Kinzler K, Vogelstein B. A model for p53induced apoptosis. Nature. 1997;389:300-305.
- Garland CF, Garland FC, Gorham ED. Rising trends in melanoma. An hypothesis concerning sunscreen effectiveness. *Annals of Epidemiology*. 1993;3:103-110.
- Naylor MF. Ozone depletion: the past 20 years. Skin Cancer Foundation Journal. 1999;17:16-17, 81-82.